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**Kylt<sup>®</sup>**

# Kylt<sup>®</sup> Viral Haemorrhagic Septicaemia Virus

Real-Time RT-PCR Detection

[www.kylt.eu](http://www.kylt.eu)



# Kylt® Viral Haemorrhagic Septicaemia Virus

## Real-Time RT-PCR Detection

### A. General

- Kylt® Viral Haemorrhagic Septicaemia Virus kits are intended for the specific detection of viral RNA of Viral Haemorrhagic Septicaemia Virus. The kits are suitable for the analysis of samples from fish, such as tissues and organs (spleen, head kidney, heart, brain), ovarian or seminal fluid, whole fish (fry), material derived from cultural processes and swabs of the aforementioned samples.
- The qualitative testing with Kylt® Viral Haemorrhagic Septicaemia Virus kits is based on a duplex Real-Time RT-PCR: In one reaction setting, the RNA target sequences for Viral Haemorrhagic Septicaemia Virus as well as for the exogenous control (Internal Control RNA (IC-RNA)) are reverse transcribed (Reverse Transcription (RT)) and amplified by respective primer pairs in the Polymerase Chain Reaction (PCR). Amplified target gene fragments are detected via fluorescently labeled probes during the PCR reaction in real-time (Real-Time PCR). The probes specific for detection of amplified Viral Haemorrhagic Septicaemia Virus as well as for the exogenous control target genes are labeled with fluorescent dye FAM and HEX, respectively, and their emitted fluorescence is separately optically measured by the Real-Time PCR thermal cycler. By means of all individual analyses in one reaction vessel per sample and the Negative Control and Positive Control per run the Viral Haemorrhagic Septicaemia Virus-specific status of a sample can be evaluated in the end. This way, results can be achieved within a few hours after sample receipt.
- These kits were developed for use by trained laboratory personnel following standardized procedures. This Direction For Use must be followed strictly.

## B. Reagents and Materials

- The following Kylt® Viral Haemorrhagic Septicaemia Virus kits are available and comprise the following reagents:

Reagent	Colour of Lid	100 Reactions Article No 31565	25 Reactions Article No 31566	Store at
2x RT-qPCR-Mix	○ transparent	4 x 280 µl	1 x 280 µl	≤ -18 °C
Detection-Mix	● violet	4 x lyophilizate (final 150 µl each)	1 x lyophilizate (final 150 µl each)	≤ -18 °C
Positive Control	● red	4 x lyophilizate (final 50 µl each)	2 x lyophilizate (final 50 µl each)	≤ -18 °C
Negative Control	● blue	2 x 1 ml	2 x 1 ml	≤ -18 °C
Kylt® Internal Control RNA (IC-RNA)	● black	2 x lyophilizate (final 500 µl)	1 x lyophilizate (final 500 µl)	≤ -18 °C

- After receipt, the components are immediately stored at ≤ -18 °C. Avoid repeated freezing and thawing of all the reagents and keep them thawed as short as possible.
- If occasional processing of few samples only is expected you may prepare appropriate aliquots of reagents before storage at ≤ -18 °C. Prepare aliquots in such a way that freeze-thaw-cycles are reduced to a maximum of three. The Negative Control can alternatively be stored at +2°C to +8°C.
- The components are to be used within the indicated shelf life (see box label). The components of different batches may not be mixed.
- Before its first use, rehydrate the Positive Control: add 50 µl of Negative Control per vial, briefly incubate at room temperature and mix thoroughly by repeated vortexing. It is recommended to generate aliquots of suitable volumes and store them at ≤ -18 °C.
- The Detection-Mix needs to be stored protected from abundant light. Do not expose to direct (sun)light. Before first use, rehydrate the lyophilized Detection-Mix: add 150 µl of the Negative Control per vial of Detection-Mix, briefly incubate at room temperature and mix by pulse-vortexing. Generate aliquots of suitable volumes and store them at ≤ -18 °C.
- Before first use rehydrate the IC-RNA by adding 500 µl of Negative Control per vial, briefly incubate at room temperature and mix by pulse-vortexing. Generate aliquots of suitable volumes and store them at ≤ -18 °C.

## C. Equipment and Reagents not included

- This detection method can be used on all commercially available Real-Time PCR thermal cyclers that detect the emitted fluorescence of the fluorescent dyes FAM and HEX (emission 520 and 550 nm, respectively). Note that default normalization option against ROX (e.g. using ABI cyclers) must be deactivated.
- Apart from the disposables, the following further devices are needed and are not included in the Kylt® Viral Haemorrhagic Septicaemia Virus kits:
  - RNA preparation kit / protocol (e.g. Kylt® RNA / DNA Purification products)
  - Table top microcentrifuge
  - Vortex
  - Micropipettes covering volumes of 1 µl to 1000 µl
  - Centrifuge for PCR tubes or plates

- Accessory Kylt® products: see chapter F “Related and Accessory Products”.
- We recommend the exclusive use of certified Nuclease-free disposables as well as powder-free protective gloves. Please wear gloves during the entire experimental procedure. Gloves need to be changed frequently, especially after spillage or suspected contaminations.

#### D. Control Reactions

- The Positive Control allows for control of the specificity and efficiency of the reagents and the reaction itself, including the performance of RT and Real-Time PCR and of the Real-Time PCR thermal cycler.
- The Negative Control allows for exclusion of contaminations. The sample testing is only valid if both, Positive and Negative Controls, are used and verified for validity in every Real-Time PCR run.
- The Internal Control RNA is added to the respective lysis buffer in standardized copy number before RNA preparation and co-purified with each sample. Add at least 5 µl of rehydrated IC-RNA per sample preparation. In case of a successful RNA preparation and the absence of RT- and Real-Time PCR inhibitors the IC-RNA can be detected in the Internal Control channel (HEX). This channel then is used to confirm true-negative test results by verifying successful RNA preparation and by excluding the presence of factors in the RNA preparation that are inhibitory to Real-Time RT-PCR.
- It is recommended to run one or more of a RNA Isolation Control (RIC) per set of RNA preparation, depending on the total number of samples processed at once. The RIC is a “mock sample” composed of the plain sterile buffer used for raw sample processing. It is randomly placed between the samples, processed like a normal sample and allows to detect potential contaminations of the reagents used (additionally to the Negative Control reaction) as well as for the detection of potential carryover contaminations between individual samples, e.g. during the RNA preparation process.

#### E. Protocol (see also „Protocol At A Glance“ at the end of this Direction For Use)

- The overall protocol of the analysis consists of the following main workflow:
  1. Sample Preparation
  2. RNA Preparation
  3. Reaction Setup, Reverse Transcription and Amplification (Real-Time RT-PCR)
  4. Data Analysis – Validity and Qualitative Result
- We recommend proceeding through the protocol without interruption to avoid potential degradation of the processed samples and reagents. If necessary, you may store the final RNA preparation at  $\leq -18\text{ °C}$  or  $\leq -70\text{ °C}$  until further processing. Avoid repeated freezing and thawing of the RNA preparations.

## 1. Sample Preparation

- We recommend pooling of at most five samples or samples from five individuals, respectively, per RNA preparation.
- Pool swabs in a sufficient volume of sterile buffer (e.g. 1 ml of Normal Saline or 0.1 x TE), let the swabs soak for an adequate period of time and finally wash out the swabs by thorough pulse-vortexing. The washed out supernatant is used for RNA preparation.
- Tissue and organ samples are homogenized thoroughly in sterile buffer (see above) and a suitable volume is used for the RNA preparation.
- Material derived from cultural processes, i.e. cell culture supernatant, can be used directly for RNA preparation.

## 2. RNA Preparation

### a) Kylt® RNA Preparation (requires Kylt® RNA / DNA Purification products (available separately))

- For detailed information on the RNA preparation process please refer to the manual of Kylt® RNA / DNA Purification products.

### b) RNA Preparation by other Methods

- Other kits or in-house methods to purify RNA may be used, as long as the quality leads to satisfactory amplification and detection of the Internal Control RNA.

## 3. Reaction Setup and Amplification (Real-Time RT-PCR)

- Before each use, briefly vortex and spin down the 2x RT-qPCR-Mix, rehydrated Detection-Mix and Negative Control.
- To determine the total number of reactions needed, count the number of samples and add two more for the Negative Control and the Positive Control (and RIC(s), if processed).
- Prepare the Master-Mix using the components listed below. A larger volume of a ready to use Master-Mix can be prepared and stored at  $\leq -18$  °C for convenient use over a longer period of time up to the expiry date given on the label. In case of frozen storage the Master-Mix should be aliquoted in such a way that freeze-thaw-cycles are reduced to a maximum of three.
- Vortex, spin down and add 16  $\mu$ l of the finalized Master-Mix to each of the PCR tubes or plate wells ("cavities").

Reagent	Volume ( $\mu$ l)	
	per Reaction	e.g. n=7
2x RT-qPCR-Mix	10 $\mu$ l	70 $\mu$ l
Detection-Mix	6 $\mu$ l	42 $\mu$ l
<b>Total Master-Mix</b>	<b>16 <math>\mu</math>l</b>	<b>112 <math>\mu</math>l</b> dispense 16 $\mu$ l per reaction
RNA (Negative Control / sample RNA / RIC(s) / Positive Control)	4.0 $\mu$ l	
<b>Total Reaction</b>	<b>20.0 <math>\mu</math>l</b>	

- Keep exposure of the 2x RT-qPCR-Mix, Detection-Mix and prepared Master-Mix to (sun)light as short as possible and return it back to appropriate storage temperature right after application. Avoid the formation of bubbles when pipetting the Master-Mix, samples and controls.
- Add 4 µl of the Negative Control to the corresponding cavity and seal it individually, if possible.
- Add 4 µl of each RNA preparation (including RIC(s), if processed) to the corresponding cavities and seal them individually, if possible.
- To minimize risk of potential cross-contaminations, 4 µl of the Positive Control are added to the corresponding cavity after all previous samples and control reactions are set up. Before each use, briefly vortex and spin down the rehydrated Positive Control (see also chapter B “Reagents and Materials”).
- If not already done, finally seal the cavities. It is recommended to briefly spin them down before the start of the Real-Time PCR run.
- Place the cavities in the Real-Time PCR thermal cycler and run the test with the Profile given below.

Kylt® Profile I				
Step No	Description	Temperature	Duration	
1	Reverse Transcription	50 °C	10 min	
2	Activation of Polymerase	95 °C	1 min	
3	Denaturation	95 °C	10 sec	} 50 cycles
4	Annealing	60 °C	75 sec	
5	Extension	72 °C	15 sec	
6	Fluorescence Detection	channel FAM and HEX		

- In the event of a combined Real-Time RT-PCR run, make sure all necessary channels are detected.
- Please follow the specified instructions of your Real-Time PCR thermal cycler as recommended by the manufacturer.

#### [4. Data Analysis – Validity and Qualitative Result](#)

##### General

- The amplification data can be processed automatically using the specific software tool of your Real-Time PCR thermal cycler. Alternatively, the threshold can be set manually considering the following directions: The threshold should cross the FAM- and HEX-curves in the linear increase of its slope (log scaling of the y-axis). By setting the threshold, the crossing point with the FAM- and HEX-curves determines the respective cycle threshold (Ct), which is negatively correlated with the initial concentration of copies of the target genes in the Real-Time RT-PCR reaction.
- Only curves with the typical exponential amplification, meaning the curve of the raw data shows a flat baseline at the beginning, followed by a clear (exponential) slope in fluorescence and possibly reaching a plateau-phase (y-axis in log scaling), should be regarded as positive.
- The actual test analysis starts with the validity check of the entire Real-Time PCR run. Afterwards, by means of the Internal Control the validity of each sample reaction and its true test result can be verified according to the Ct-value of the Internal Control channel (HEX). Finally, the Viral Haemorrhagic Septicaemia Virus-specific status of each sample is analyzed (FAM).

## Test Evaluation - Control Reactions

- The **Real-Time PCR test run** is only **valid** if the curves of the control reactions can be evaluated as follows:

Control Reactions	Channel	
	HEX	FAM
Negative Control	negative	negative
Positive Control	negative	<b>positive</b>

- For a valid test the FAM-Ct-value of the Positive Control has to be  $> 15$  and  $\leq 35$
- Depending on the cycler and consumables used strong signals in the FAM-channel of the Positive Controls may lead to weak background signals in the HEX-channel.
- If one or more of a RNA Isolation Control (RIC(s)) is processed, its FAM-curves must be negative and its HEX-curve must be positive.

## Test Evaluation - Samples

Target	Channel	Signal		
Internal Control	HEX	<b>positive</b>	positive / negative	negative
Viral Haemorrhagic Septicaemia Virus	FAM	negative	<b>positive</b>	negative
<b>The sample is Viral Haemorrhagic Septicaemia Virus</b>		<b>negative</b>	<b>positive</b>	<b>inhibited</b>

- A **sample is negative for Viral Haemorrhagic Septicaemia Virus**, if its HEX-curve is positive ( $Ct \leq 40$ ), but its FAM-curve is negative.
- A **sample is positive for Viral Haemorrhagic Septicaemia Virus**, if its FAM-curve is positive ( $Ct \leq 42$ ), independent of the HEX-curve.
- A **sample is inhibited**, if neither the HEX-curve nor the FAM-curve is positive.
- **Recommendation:** In the case of an inhibited sample you may repeat the test by using e.g. 1:4 dilution of the respective RNA preparation. The Negative Control is used as the diluting agent. Preferably, the entire RNA preparation process is repeated using Kylt® RNA/DNA Purification products or appropriate alternative.
- Convenient and reliable sample data entry, Real-Time PCR start, final qualitative analysis and documentation can be conducted with the Kylt® Software, please inquire.

## F. Related and Accessory Products

Product	Article No	Content	Description
Kylt® RNA / DNA Purification	31315	50	Combined RNA and DNA purification from all relevant sample matrices
Kylt® RNA / DNA Purification HTP	31826	4 x 96	Combined, magnetic beads-based purification of RNA and DNA from all relevant sample matrices, suitable for automated high throughput processing
Kylt® Purifier	31436	1 unit	Purification system for magnetic bead based kits. Up to 96 samples are processed in under 30 minutes. Intended for high-throughput laboratories.
Kylt® Purifier 48	31436	1 unit	Purification system for magnetic bead based kits. Up to 48 samples are processed in under 30 minutes. Intended for low to medium throughput laboratories.
Kylt® Purifier Spin Tips	31434	5 Sets	Plate with 96 separate spin tips, used by the Kylt® Purifier to mix the well contents by stirring. Sufficient for 480 samples.
Kylt® Purifier Plates	31435	20 Plates	Plates to be used for the several reactions and reagents during automated nucleic acid purification. Sufficient for 320 to 480 samples (depending on device and protocol).

## G. Ordering information

For a fast and efficient service please send your order to [orders@kylt.eu](mailto:orders@kylt.eu) and please provide the following information:

- Delivery and Invoice address
- Purchaser contact telephone number
- End user name and telephone number (if different)
- Purchase order number
- Product name and catalogue number
- Quantity and size of products
- Indicate if your account is VAT exempt



Production:

AniCon Labor GmbH | Muehlenstr. 13 | D-49685 Hoeltinghausen | Germany | [www.kylt.eu](http://www.kylt.eu) | [info@kylt.eu](mailto:info@kylt.eu)

Development, manufacturing and distribution of Kylt® *In-Vitro* Diagnostica is certified according to ISO 9001:2015.

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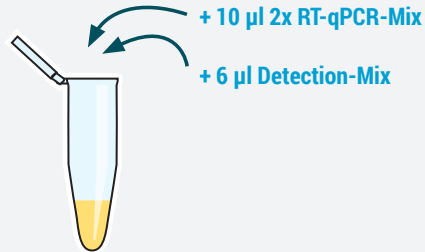


# PROTOCOL AT A GLANCE

## Real-Time RT-PCR Setup

1

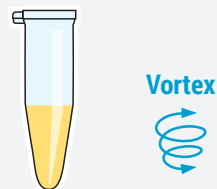
Prepare a Master-Mix\*



\* please refer to chapter 3

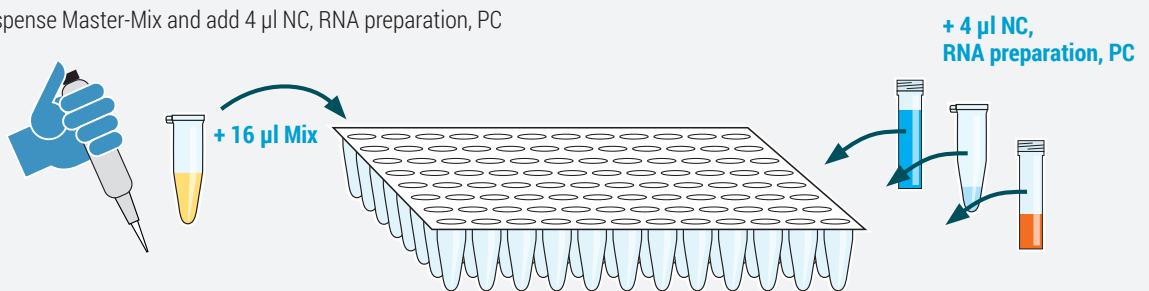
2

Pulse-vortex and spin down



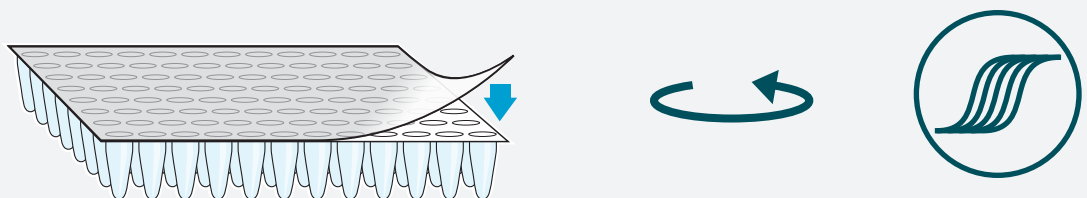
3

Dispense Master-Mix and add 4 µl NC, RNA preparation, PC



4

Seal cavities, spin down (recommended), and start cycler



5

Analysis

