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Kynt[®]

Kynt[®] E.coli iucD, pic, cnf1, eae

Real-Time PCR Detection

www.kynt.eu

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A. General

- Kylt® E.coli iucD, pic, cnf1, eae kits are intended for the specific detection of bacterial DNA of *Escherichia coli* virulence factors iucD, pic, cnf1 and eae. The kits are only to be used with pure colony material / isolates derived from cultural processes.
- The qualitative testing with Kylt® E. coli iucD, pic, cnf1, eae is based on a multiplex Real-Time PCR: In one reaction setting, the target genes for *E. coli* virulence factors iucD, pic, cnf1 and eae are amplified in parallel by respective primer pairs in the Polymerase Chain Reaction (PCR). Amplified target gene fragments are detected via fluorescently labeled probes during the PCR reaction in real-time (Real-Time PCR). The probes specific for detection of amplified *E. coli* iucD, pic, cnf1, eae target genes are labeled with fluorescent dyes FAM, Cy5, HEX, and TXR, respectively, and their emitted fluorescence is separately optically measured by the Real-Time PCR thermal cycler. By means of all four individual analyses in one reaction vessel per sample and the Negative Control and Positive Control per run the *E. coli* iucD, pic, cnf1, eae-specific status of a sample can be evaluated in the end. This way, results can be achieved within a few hours after sample receipt.
- These kits were developed for use by trained laboratory personnel following standardized procedures. This Direction For Use must be followed strictly.

B. Reagents and Materials

- The following Kylt® E.coli iucD, pic, cnf1, eae kits are available and comprise the following reagents:

Reagent	Colour of Lid	100 Reactions	25 Reactions	Store at
		Article No 31557	Article No 31558	
Reaction-Mix	● green	4 x 450 µl	1 x 450 µl	≤ -18 °C
Positive Control	● red	4 x lyophilizate (final 50 µl each)	2 x lyophilizate (final 50 µl each)	≤ -18 °C
Negative Control	● blue	1 x 1 ml	1 x 1 ml	≤ -18 °C

- After receipt, the components are immediately stored at ≤ -18 °C. Avoid repeated freezing and thawing of all the reagents and keep them thawed as short as possible. If occasional processing of few samples only is expected you may prepare appropriate aliquots of reagents before storage at ≤ -18 °C. Prepare aliquots in such a way that freeze-thaw-cycles are reduced to a maximum of three. The Negative Control can alternatively be stored at +2°C to +8°C.
- The components are to be used within the indicated shelf life (see box label). The components of different batches may not be mixed.
- Before its first use, rehydrate the Positive Control: add 50 µl of Negative Control per vial, briefly incubate at room temperature and mix thoroughly by repeated vortexing. It is recommended to generate aliquots of suitable volumes and store them at ≤ -18 °C.
- The Reaction-Mix needs to be stored protected from abundant light. Do not expose to direct (sun)light.

C. Equipment and Reagents not included

- This detection method can be used on all commercially available Real-Time PCR thermal cyclers that detect the emitted fluorescence of the fluorescent dyes FAM, Cy5, HEX and TXR (emission 520, 670, 550 and 620 nm, respectively). Note that default normalization option against ROX (e.g. using ABI cyclers) must be deactivated.
- Apart from the disposables, the following further devices are needed and are not included in the Kylt® E. coli iucD, pic, cnf1, eae kits:
 - DNA preparation kit / protocol (e.g. Kylt® DNA Extraction-Mix II or Kylt® RNA / DNA Purification products)
 - Table top microcentrifuge
 - Vortex
 - Micropipettes covering volumes of 1 µl to 1000 µl
 - Centrifuge for PCR tubes or plates
- Accessory Kylt® products: see chapter F “Related and Accessory Products”.
- We recommend the exclusive use of certified Nuclease-free disposables as well as powder-free protective gloves. Please wear gloves during the entire experimental procedure. Gloves need to be changed frequently, especially after spillage or suspected contaminations.

D. Control Reactions

- The Positive Control allows for control of the specificity and efficiency of the reagents and the reaction itself, including the performance of the Real-Time PCR and of the Real-Time PCR thermal cycler.
- The Negative Control allows for exclusion of contaminations. The sample testing is only valid if both, Positive and Negative Controls, are used and verified for validity in every Real-Time PCR run.

E. Protocol *(see also „Protocol At A Glance“ at the end of this Direction For Use)*

- The overall protocol of the analysis consists of the following main workflow:
 1. Sample Preparation
 2. DNA Preparation
 3. Reaction Setup and Amplification (Real-Time PCR)
 4. Data Analysis – Validity and Qualitative Result
- We recommend proceeding through the protocol without interruption to avoid potential degradation of the processed samples and reagents. If necessary, you may store the final DNA preparation at ≤ -18 °C until further processing. Avoid repeated freezing and thawing of the DNA preparations.

1. Sample Preparation

- Material derived from cultural processes, i.e. colony material, is directly transferred into respective tubes for Kylt® DNA Extraction (please refer to 2 "DNA Preparation"), such as conical screw cap tube; therefore a little amount of a single colony is picked with a sterile loop wire or sterile pipette tip and transferred to the tube.

2. DNA Preparation

a) Kylt® DNA Extraction (requires Kylt® DNA Extraction-Mix II)

- For detailed information, please refer to the Direction For Use of Kylt® DNA Extraction-Mix II.

b) Kylt® RNA/DNA Purification products

- All kinds of sample matrices, including pure isolates may be processed with Kylt® RNA/DNA Purification products (please refer to chapter F "Related Products").
- For detailed information on the DNA preparation process, please refer to the respective Direction For Use.

c) Alternative methods

- All kinds of sample matrices, including pure isolates may be processed with appropriate DNA preparation kits or appropriate in-house methods.
- For detailed information on the DNA preparation process, please refer to the Direction For Use or Standard Operating Procedure of the specific kit or in-house method, respectively.

3. Reaction Setup and Amplification (Real-Time PCR)

- Before each use, briefly vortex and spin down the Reaction-Mix and Negative Control.
- To determine the total number of reactions needed, count the number of samples and add two more for the Negative Control and the Positive Control.
- The Reaction-Mix is ready-to-use, add 16 µl to each of the PCR tubes or plate wells (“cavities”).
- Keep exposure of the Reaction-Mix to (sun)light as short as possible and return it back to appropriate storage temperature right after application. Avoid the formation of bubbles when pipetting samples and controls.
- Add 4 µl of the Negative Control to the corresponding cavity and seal it individually, if possible.
- Add 4 µl of each DNA preparation to the corresponding cavities and seal them individually, if possible.
- To minimize risk of potential cross-contaminations, 4 µl of the Positive Control are added to the corresponding cavity after all previous samples and control reactions are set up. Before each use, briefly vortex and spin down the rehydrated Positive Control (see also chapter B “Reagents and Materials”).
- If not already done, finally seal the cavities. It is recommended to briefly spin them down before the start of the Real-Time PCR run.
- Place the cavities in the Real-Time PCR thermal cycler and run the test with KyIt® Profile II as given below.

KyIt® Profile II				
Step No	Description	Temperature	Duration	
1	Activation of Polymerase	95 °C	10 min	
2	Denaturation	95 °C	15 sec	} 42 cycles
3	Annealing & Extension	60 °C	1 min	
4	Fluorescence Detection	channels FAM, Cy5, HEX and TXR		

- KyIt® Profile II allows for combined run of this and most other KyIt® qPCR detection methods.
- Alternatively, the KyIt® Profile I given below can be applied. KyIt® Profile I allows for combined run of this and most other KyIt® qPCR detection methods as well as KyIt® RT-qPCR detection products that need Reverse Transcription, such as those for detection of viral RNA.

KyIt® Profile I				
Step No	Description	Temperature	Duration	
1	Reverse Transcription	50 °C	10 min	
2	Activation of Polymerase	95 °C	1 min	
3	Denaturation	95 °C	10 sec	} 42 cycles
4	Annealing & Extension	60 °C	1 min	
5	Fluorescence Detection	channels FAM, Cy5, HEX and TXR		

- In the event of a combined Real-Time (RT)-PCR run, make sure all necessary channels are detected.
- Please follow the specified instructions of your Real-Time PCR thermal cycler as recommended by the manufacturer.

4. Data Analysis – Validity and Qualitative Result

General

- The amplification data can be processed automatically using the specific software tool of your Real-Time PCR thermal cycler. Alternatively, the threshold can be set manually considering the following directions: The threshold should cross the FAM, Cy5, HEX and TXR-curves in the linear increase of their slope (log scaling of the y-axis). By setting the threshold, the crossing points with the FAM, Cy5, HEX and TXR-curves determine the respective cycle threshold (Ct), which is negatively correlated with the initial concentration of copies of the target genes in the Real-Time PCR reaction.
- Only curves with the typical exponential amplification, meaning the curve of the raw data shows a flat baseline at the beginning, followed by a clear (exponential) slope in fluorescence and possibly reaching a plateau-phase (y-axis in log scaling), should be regarded as positive.
- The actual test analysis starts with the validity check of the entire Real-Time PCR run. Finally, the *E. coli* iucD, pic, cnf1, eae-specific status of each sample is analyzed (FAM, Cy5, HEX and TXR).

Test Evaluation - Control Reactions

- The **Real-Time PCR test run** is only **valid** if the curves of the control reactions can be evaluated as follows:

Control Reactions	Channel			
	FAM	Cy5	HEX	TXR
Negative Control	negative	negative	negative	negative
Positive Control	positive	positive	positive	positive

- For a valid test the Ct-values (FAM, Cy5, HEX and TXR) of the Positive Control have to be > 15 and ≤ 30.
- *E. coli* iucD, pic, cnf1, eae can only be analyzed in *E. coli* positive samples or isolates.

Test Evaluation

Target	Channel					
iucD	FAM	positive	negative	negative	negative	negative
pic	Cy5	negative	positive	negative	negative	negative
cnf1	HEX	negative	negative	positive	negative	negative
eae	TXR	negative	negative	negative	positive	negative
The sample is <i>E. coli</i> iucD		positive	negative	negative	negative	<i>E. coli</i> iucD, pic, cnf1 and eae not detectable*
The sample is <i>E. coli</i> pic		negative	positive	negative	negative	
The sample is <i>E. coli</i> cnf1		negative	negative	positive	negative	
The sample is <i>E. coli</i> eae		negative	negative	negative	positive	

* This result may also indicate PCR inhibition. To address this it is recommended to test the respective sample with another assay containing an internal amplification control (e.g. Kylt® *E. coli* papC, escV, cdtB)

- A **sample is positive for *E. coli* iucD**, if its FAM-curve is positive (Ct ≤ 30), independent of the other curves.
- A **sample is positive for *E. coli* pic**, if its Cy5-curve is positive (Ct ≤ 30), independent of the other curves.
- A **sample is positive for *E. coli* cnf1**, if its HEX-curve is positive (Ct ≤ 30), independent of the other curves.
- A **sample is positive for *E. coli* eae**, if its TXR-curve is positive (Ct ≤ 30), independent of the other curves.
- A **sample is negative for *E. coli* iucD, pic, cnf1 and eae**, if its FAM-, Cy5-, HEX- and TXR-curves are negative.
- For the Kylt® *E. coli* iucD, pic, cnf1, eae Real-Time PCR Detection cut-off values have to be set for the iucD, pic, cnf1 and eae specific channels. Only results of Ct-value below Ct 30 for the specific channels FAM, Cy5, HEX and TXR are to be considered as valid and positive.
- The Ct cut-off has no impact on the sensitivity because the sample material for the Kylt® *E. coli* iucD, pic, cnf1, eae Real-Time PCR Detection is colony material derived from cultural processes and therefore gives strong positive signals.
- Convenient and reliable sample data entry, Real-Time PCR start, final qualitative analysis and documentation can be conducted with the Kylt® Software, please inquire.

F. Related and Accessory Products

Product	Article No	Reactions	Description
Kylt® DNA Extraction-Mix II	31398	100	Simplified and economic DNA extraction
Kylt® RNA / DNA Purification	31315	50	Combined RNA and DNA purification from veterinary samples (spin-column based).
Kylt® RNA / DNA Purification HTP	31826	4 x 96	Magnetic bead based combined RNA and DNA purification kit for veterinary diagnostic samples. Suitable for Kylt® Purifier and Kylt® Purifier 48.
Kylt® Purifier	31436	1 unit	Purification system for magnetic bead based kits. Up to 96 samples are processed in under 30 minutes. Intended for high-throughput laboratories.
Kylt® Purifier 48	31436	1 unit	Purification system for magnetic bead based kits. Up to 48 samples are processed in under 30 minutes. Intended for low to medium throughput laboratories.
Kylt® Purifier Spin Tips	31434	5 Sets	Plate with 96 separate spin tips, used by the Kylt® Purifier to mix the well contents by stirring. Sufficient for 480 samples.
Kylt® Purifier Plates	31435	20 Plates	Plates to be used for the several reactions and reagents during automated nucleic acid purification. Sufficient for 320 to 480 samples (depending on device and protocol) .

G. Ordering information

For a fast and efficient service please send your order to orders@kylt.eu and please provide the following information:

- Delivery address
- Invoice address
- Purchaser contact telephone number
- End user name and telephone number (if different)
- Purchase order number
- Product name and catalogue number
- Quantity and size of products
- Indicate if your account is VAT exempt

Production:

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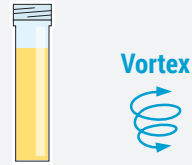


PROTOCOL AT A GLANCE

Real-Time PCR Setup

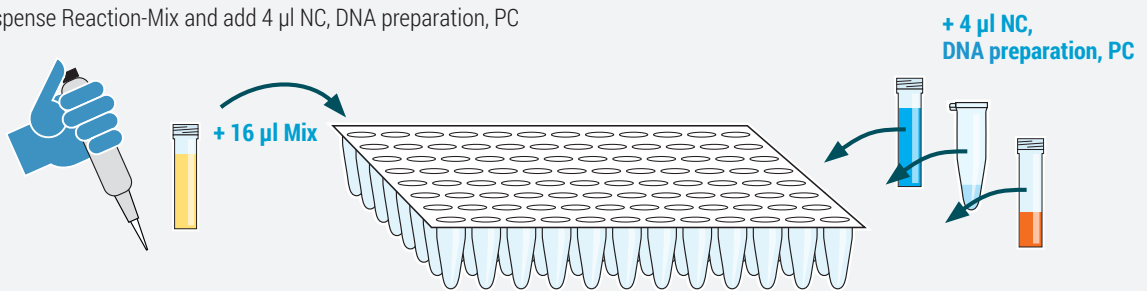
1

Pulse-vortex and spin down



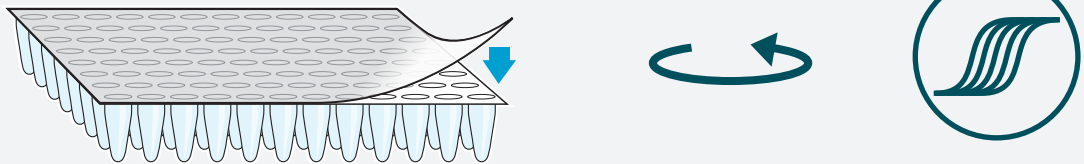
2

Dispense Reaction-Mix and add 4 µl NC, DNA preparation, PC



3

Seal cavities, spin down (recommended), and start cycler



4

Analysis

