



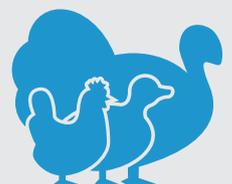
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Diagnostics only.

Kylt[®]

Kylt[®] IBV-Variant Arkansas

Real-Time RT-PCR Detection

www.kylt.eu



Kylt® IBV-Variant Arkansas

Real-Time RT-PCR Detection

A. General

- Kylt® IBV-Variant Arkansas products are intended for the specific detection of viral RNA of Infectious Bronchitis Virus (IBV) variant Arkansas. The products are suitable for the analysis of samples from birds such as tissues and organs (e.g. trachea, lung, kidney, cecal tonsils), swab samples of the aforementioned tissues and organs and samples from cultural processes with the aforementioned sample material.
- The qualitative testing with Kylt® IBV-Variant Arkansas products is based on a Real-Time RT-PCR: In one reaction setting, the target gene for IBV-Variant Arkansas is reverse transcribed (Reverse Transcription (RT)) and amplified by respective primer pairs in the Polymerase Chain Reaction (PCR). Amplified target gene fragments are detected via fluorescently labeled probes during the PCR reaction in real-time (Real-Time PCR). The probe specific for detection of amplified IBV-Variant Arkansas target gene is labeled with fluorescent dye Cy5, and its emitted fluorescence is optically measured by the Real-Time PCR thermal cycler. By means of this analysis in one reaction vessel per sample and the Negative Control and Positive Control per run the IBV-Variant Arkansas-specific status of a sample can be evaluated in the end. This way, results can be achieved within a few hours after sample receipt.
- It is recommended to initially screen samples with Kylt® IB-aCoV Real-Time RT-PCR Detection to rule out negative samples and to assess potential PCR-inhibitors in the sample (see chapter F "Related and Accessory Products").
- These products were developed for use by trained laboratory personnel following standardized procedures. This Direction For Use must be followed strictly.

B. Reagents and Materials

- The following Kylt® IBV-Variant Arkansas products are available and comprise the following reagents:

Reagent	Colour of Lid	Quantity in Kit with 100 / 25 Reactions		Store at
		Article No 31088 / 31089	Article No# 31465 / 31466	
2x RT-qPCR-Mix	○ transparent	4 x / 1 x 280 µl	-/-	≤ -18 °C
Detection-Mix	● yellow	4 x / 1 x lyophilizate (final 150 µl each)	4 x / 1 x lyophilizate (final 150 µl each)	≤ -18 °C
Positive Control	● red	4 x / 2 x lyophilizate (final 50 µl each)	4 x / 2 x lyophilizate (final 50 µl each)	≤ -18 °C
Negative Control	● blue	1 x 1 ml	1 x 1 ml	≤ -18 °C

To conduct an analysis for IBV-Variant Arkansas an additional, at least 2x concentrated RT-qPCR-Mix is required, please refer to chapter C. "Equipment and Reagents not included" or inquire for recommendations.

- After receipt, the components are immediately stored at ≤ -18 °C. Avoid repeated freezing and thawing of all the reagents and keep them thawed as short as possible. If occasional processing of few samples only is expected you may prepare appropriate aliquots of reagents before storage at ≤ -18 °C. Prepare aliquots in such a way that freeze-thaw-cycles are reduced to a maximum of three. The Negative Control can alternatively be stored at +2°C to +8°C.
- The components are to be used within the indicated shelf life (see label on the outer packing). The components of different batches may not be mixed.
- Before its first use, rehydrate the Positive Control: add 50 µl of Negative Control per vial, briefly incubate at room temperature and mix thoroughly by repeated vortexing. It is recommended to generate aliquots of suitable volumes and store them at ≤ -18 °C.
- The Detection-Mix needs to be stored protected from abundant light. Do not expose to direct (sun) light. Before first use, rehydrate the lyophilized Detection-Mix: add 150 µl of the Negative Control per vial of Detection-Mix, briefly incubate at room temperature and mix by pulse-vortexing. Generate aliquots of suitable volumes and store them at ≤ -18 °C.

C. Equipment and Reagents not included

- This detection method can be used on all commercially available Real-Time PCR thermal cyclers that detect the emitted fluorescence of the fluorescent dye Cy5 (emission 670 nm). Note that default normalization option against ROX (e.g. using ABI cyclers) must be deactivated.
- Apart from the disposables, the following further devices are needed and are not included in the Kylt® IBV-Variant Arkansas products:
 - RNA preparation kit / protocol (e.g. Kylt® RNA / DNA Purification)
 - Table top microcentrifuge
 - Vortex
 - Micropipettes covering volumes of 1 µl to 1000 µl
 - Centrifuge for PCR tubes or plates
- For Kylt® products supplied with Detection-Mix only, at least 2x concentrated RT-qPCR-Mix is needed (e.g. BCD 2x RT-qPCR-Mix).

- Accessory Kylt® products: see chapter F “Related and Accessory Products”.
- We recommend the exclusive use of certified Nuclease-free disposables as well as powder-free protective gloves. Please wear gloves during the entire experimental procedure. Gloves need to be changed frequently, especially after spillage or suspected contaminations.

D. Control Reactions

- The Positive Control allows for control of the specificity and efficiency of the reagents and the reaction itself, including the performance of RT and Real-Time PCR and of the Real-Time PCR thermal cycler.
- The Negative Control allows for exclusion of contaminations. The sample testing is only valid if both, Positive and Negative Controls, are used and verified for validity in every Real-Time PCR run.
- It is recommended to run one or more of a RNA Isolation Control (RIC) per set of RNA preparation, depending on the total number of samples processed at once. The RIC is a “mock sample” composed of the plain sterile buffer used for raw sample processing. It is randomly placed between the samples, processed like a normal sample and allows to detect potential contaminations of the reagents used (additionally to the Negative Control reaction) as well as for the detection of potential carryover contaminations between individual samples, e.g. during the RNA preparation process.

E. Protocol *(see also „Protocol At A Glance“ at the end of this Direction For Use)*

- The overall protocol of the analysis consists of the following main workflow:
 1. Sample Preparation
 2. RNA Preparation
 3. Reaction Setup, Reverse Transcription and Amplification (Real-Time RT-PCR)
 4. Data Analysis – Validity and Qualitative Result
- We recommend proceeding through the protocol without interruption to avoid potential degradation of the processed samples and reagents. If necessary, you may store the final RNA preparation at $\leq -18\text{ °C}$ or $\leq -70\text{ °C}$ until further processing. Avoid repeated freezing and thawing of the RNA preparations.
- It is recommended to initially screen samples with the Kylt® IB-aCoV Real-Time RT-PCR Detection (see chapter F “Related and Accessory Products”).

1. Sample Preparation

- We recommend pooling of at most five samples or samples from five individuals, respectively, per RNA preparation.
- Pool swabs in a sufficient volume of sterile buffer (e.g. 1 ml of Normal Saline or 0.1 x TE), let the swabs soak for an adequate period of time and finally wash out the swabs by thorough pulse-vortexing. The washed out supernatant is used for RNA preparation.
- Tissue and organ samples are homogenized thoroughly in sterile buffer (see above) and a suitable volume is used for the RNA preparation.
- Material derived from cultural processes, i.e. cell culture supernatant or allantoic fluid, can be used directly for RNA preparation.

2. RNA Preparation

a) Kylt® RNA Preparation (requires Kylt® RNA / DNA Purification)

- For detailed information on the RNA preparation process please refer to the manual of Kylt® RNA / DNA Purification.

b) RNA Preparation by other Methods

- Other kits or in-house methods to purify RNA may be used, as long as the quality leads to satisfactory amplification and detection of the Internal Control RNA.

3. Reaction Setup and Amplification (Real-Time RT-PCR)

- Before each use, briefly vortex and spin down the 2x RT-qPCR-Mix, rehydrated Detection-Mix and Negative Control.
- To determine the total number of reactions needed, count the number of samples and add two more for the Negative Control and the Positive Control (and RIC(s), if processed).
- Prepare the Master-Mix using the components listed below. It is strongly recommended to prepare a Master-Mix consisting of 2x RT-qPCR-Mix (e.g. BCD 2x RT-qPCR-Mix) and Detection-Mix, at least per PCR setup.
- When calculating volumes for preparation of the Master-Mix consider the respective concentration of the RT-qPCR-Mix used. For more information about the RT-qPCR-Mix please refer to the Direction For Use of the respective manufacturer.
- Alternatively, when using the included 2x RT-qPCR Mix or the recommended BCD 2x RT-qPCR Mix a larger volume of a ready to use Master-Mix can be prepared and stored at ≤ -18 °C for convenient use over a longer period of time up to the expiry date given on the label. In case of frozen storage the Master-Mix should be aliquoted in such a way that freeze-thaw-cycles are reduced to a maximum of three.
- Vortex, spin down and add 16 μ l of the finalized Master-Mix to each of the PCR tubes or plate wells ("cavities").

Example Calculation for 2x concentrated RT-qPCR-Mix:	Volume (μ l)	
	per Reaction	e.g. n=7
2x RT-qPCR-Mix (e.g. BCD 2x RT-qPCR-Mix)	10 μ l	70 μ l
Detection-Mix	6 μ l	42 μ l
Total Master-Mix	16 μl	112 μl dispense 16 μ l per reaction
RNA (Negative Control / sample RNA / RIC(s) / Positive Control)	4.0 μ l	
Total Reaction	20.0 μl	

- Keep exposure of the 2x RT-qPCR-Mix, Detection-Mix and prepared Master-Mix to (sun)light as short as possible and return it back to appropriate storage temperature right after application. Avoid the formation of bubbles when pipetting the Master-Mix, samples and controls.
- Add 4 μ l of the Negative Control to the corresponding cavity and seal it individually, if possible.
- Add 4 μ l of each RNA preparation (including RIC(s), if processed) to the corresponding cavities and seal them individually, if possible.

- To minimize risk of potential cross-contaminations, 4 µl of the Positive Control are added to the corresponding cavity after all previous samples and control reactions are set up. Before each use, briefly vortex and spin down the rehydrated Positive Control (see also chapter B “Reagents and Materials”).
- If not already done, finally seal the cavities. It is recommended to briefly spin them down before the start of the Real-Time PCR run.
- Place the cavities in the Real-Time PCR thermal cycler and run the test with Kylt® Profile III as given below.
- In case Kylt® products supplied with Detection-Mix are used in combination with the BCD 2x RT-qPCR Mix, preferably the given Kylt® Profile III is applied. For other RT-qPCR Mixes, please refer to the respective Direction For Use, check for appropriateness and adapt duration or temperature of individual steps, if necessary.

Kylt® Profile III				
Step No	Description	Temperature	Duration	
1	Reverse Transcription	50 °C	10 min	
2	Activation of Polymerase	95 °C	1 min	
3	Denaturation	95 °C	10 sec	} 42 cycles
4	Annealing & Extension	55 °C	1 min	
5	Fluorescence Detection	channel Cy5		

- Kylt® Profile III allows for combined run of the Kylt® RT-qPCR detection products for IBV variants and IBDV.
- In the event of a combined Real-Time RT-PCR run, make sure all necessary channels are detected.
- Please follow the specified instructions of your Real-Time PCR thermal cycler as recommended by the manufacturer.

4. Data Analysis – Validity and Qualitative Result

General

- The amplification data can be processed automatically using the specific software tool of your Real-Time PCR thermal cycler. Alternatively, the threshold can be set manually considering the following directions: The threshold should cross the Cy5-curve in the linear increase of their slope (log scaling of the y-axis). By setting the threshold, the crossing point with the Cy5-curve determines the respective cycle threshold (Ct), which is negatively correlated with the initial concentration of copies of the target genes in the Real-Time RT-PCR reaction.
- Only curves with the typical exponential amplification, meaning the curve of the raw data shows a flat baseline at the beginning, followed by a clear (exponential) slope in fluorescence and possibly reaching a plateau-phase (y-axis in log scaling), should be regarded as positive.

Test Evaluation

- The **Real-Time RT-PCR test run** is only **valid** if the Cy5-curve of the Negative Control is negative and the Cy5-curve of the Positive Control is positive. For a valid test the Cy5-Ct-value of the Positive Control has to be > 15 and ≤ 35.
- If one or more of a RNA Isolation Control (RIC(s)) is processed, its Cy5-curve must be negative.

Target	Channel	Signal	
IBV-Variant Arkansas	Cy5	negative	positive
The sample is IBV-Variant Arkansas		negative	positive

- A **sample is negative for IBV-Variant Arkansas** if its Cy5-curve is negative.
- A **sample is positive for IBV-Variant Arkansas** if its Cy5-curve is positive (Ct ≤ 42).
- Convenient and reliable sample data entry, Real-Time PCR start, final qualitative analysis and documentation can be conducted with the Kylt® Software, please inquire.

F. Related and Accessory Products

Product	Article No	Reactions	Description
Kylt® RNA / DNA Purification	31314 / 31315	250 / 50	Combined RNA and DNA purification from veterinary samples
Kylt® IB-aCoV	31193 / 31194	100 / 25	Screening for Infectious Bronchitis Virus and further avian Coronaviruses
BCD 2x RT-qPCR-Mix	31128	100	2-fold concentrated Real-Time RT-PCR mix

Production:

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Development, manufacturing and distribution of Kylt® *In-Vitro* Diagnostica is certified according to ISO 9001:2015.

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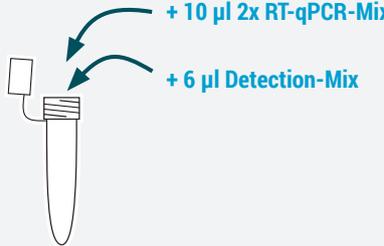
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PROTOCOL AT A GLANCE

Real-Time RT-PCR Setup

1 KyIt® Products supplied with
▼ Detection Mix & 2x RT-qPCR-Mix

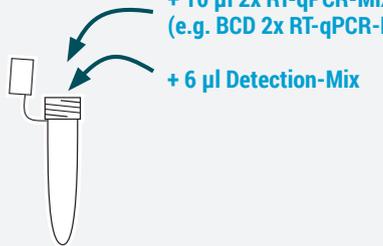
Prepare a Master-Mix*



* please refer to chapter 3

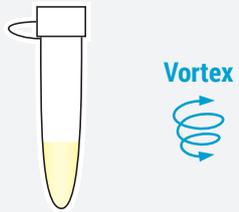
KyIt® Products supplied with
▼ Detection Mix

Prepare a Master-Mix*

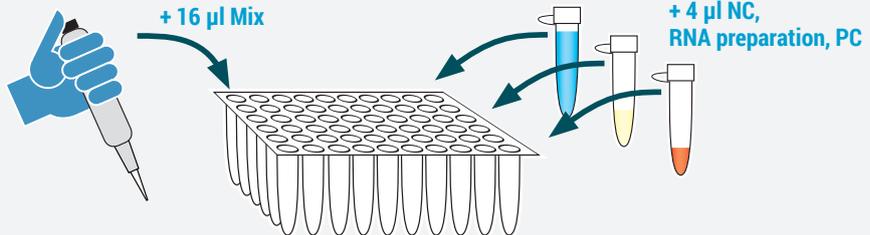


* please refer to chapter 3

2 Pulse-vortex and spin down



3 Dispense Master-Mix and add 4 µl NC, RNA preparation, PC



4 Seal cavities, spin down (recommended), and start cycler



5 Analysis

