C. Protocol

1. Sample Preparation

- **Recommended:** Transfer at least 3 ml of supernatant of the enrichment to a sterile screw cap tube using sterile transfer pipettes, and discard the Stomacher bag. The tube is used as an intermediary container as well as store the sample for potential cultivation of Salmonellae until testing is complete.
- **Attention:** Mixing of the pre-enrichment after incubation by shaking or any other agitation must be avoided! Avoid transferring solid or greasy debris. An aliquot should be taken directly below the surface, avoiding floating particles.
- An aliquot of 500 µl per sample is transferred to each used well a Kylt® Purifier Plate (apply label: "Bind").

2. Plate Preparation

- Add 500 µl Wash Solution to each used well of a Kylt® Purifier Plate (apply label: "Wash").
- Add 500 µl Ethanol Wash to each used well of a Kylt® Purifier Plate (apply label: "EtOH").
- Add 100 µl Elution Buffer to each used well of a Kylt® Purifier Plate (apply label: "Elution").

3. Binding

- Resuspend Binding Reagent by inverting the bottle several times to obtain a homogenous suspension.
- Transfer the necessary volume to a pipetting reservoir.
- Using an electronic multi-channel pipette, transfer 750 µl of homogenous Binding Reagent to each well and mix contents twice.

4. Starting Kylt® Purifier

- Power on Kylt® Purifier.
- Choose "Run", choose "Kylt® Salmonella" protocol.
- Follow instructions and load machine as prompted, column 1 of each plate should point to the inside of the machine.
- Start purification.

5.Unloading Kylt® Purifier

- The protocol is finished in about 25 minutes.
- The first accessible plate contains the eluates.
- The eluates can be used directly in the Kylt® Salmonella spp. Real-Time PCR.
- Transfer eluates to another plate for storage, if needed. Seal with foil seal.
- Unload Kylt® Purifier by turning the table with the buttons, discard plates accordingly.
- Clean surfaces and perform UV-decontamination if necessary.